



**GeneQuery™ Human Hepatic Steatosis qPCR Array Kit
(GQH-HST)
Catalog #GK045**

Product Description

ScienCell's GeneQuery™ Human Hepatic Steatosis qPCR Array Kit (GQH-HST) is designed to facilitate gene expression profiling of 88 key genes involved in hepatic steatosis pathogenesis. Hepatic steatosis, also known as fatty liver disease, refers to the accumulation of excessive triglycerides and other lipids in hepatocytes, mainly due to defective fatty acid metabolism. The most common causes of hepatic steatosis include hepatic insulin resistance, alcoholism, obesity or imbalance between energy intake and consumption, and genetic inheritance. Brief examples of how included genes may be grouped are shown below:

- **Insulin signaling pathways:** SOCS3, PPARGC1A, AKT1, MTOR, GSK3B, INSR, IRS1, FOXO1, PIK3R1, SOCS3, SREBF1
- **Cholesterol metabolism:** APOC3, APOA1, APOB, APOE, LDLR, PPARG, SREBF1, SREBF2, NR1H3, CYP2E1
- **Carbohydrate metabolism:** G6PC, G6PD, ACLY, PCK2, PDK4, RBP4, GSK3B
- **Beta-oxidation:** CPT1A, CPT2, FABP1, ACADL, ACOX1, AKT1, CD36, MTOR
- **Adipokine signaling pathways:** PPARGC1A, MTOR, LEPR, IRS1, ADIPOR1, ADIPOR2, NFKB1, SLC2A1, SLC2A4, TNF
- **Hepatotoxicity steatosis:** CD36, FASN, LPL, SCD, PPARA, SREBF1
- **Genes implicated in**
 - **Type II diabetes:** IRS1, INSR, MAPK8, SLC2A2, SLC2A4, XBP1, TNF, PKLR, PNPLA3
 - **Non-alcoholic fatty liver disease:** PNPLA3, SQSTM1, INS, GPT, ADIPOQ, TNF, SLC17A5, LEP, KRT18
 - **Reye's syndrome:** ACADM, OTC, HMGCL, HADHA, OAT, ASS1, HMGCR, CES1
 - **Lipodystrophy:** LMNA, LEP, IRS4, PDE3B, PTRF, GFPT1

GeneQuery™ qPCR array kits are qPCR ready in a 96-well plate format, with each well containing one primer set that can specifically recognize and efficiently amplify a target gene's cDNA. The carefully designed primers ensure that: (i) the optimal annealing temperature in qPCR analysis is 65°C (with 2 mM Mg²⁺, and no DMSO); (ii) the primer set recognizes all known transcript variants of target gene, unless otherwise indicated; and (iii) only one gene is amplified. Each primer set has been validated by qPCR with melt curve analysis, and gel electrophoresis.

GeneQuery™ qPCR Array Kit Controls

Each GeneQuery™ plate contains eight controls (Figure 1).

- Five target housekeeping genes (ACTB, GAPDH, LDHA, NONO, and PPIH), which enable normalization of data.

- The Genomic DNA (gDNA) Control (GDC) detects possible gDNA contamination in the cDNA samples. It contains a primer set targeting a non-transcribed region of the genome.
- Positive PCR Control (PPC) tests whether samples contain inhibitors or other factors that may negatively affect gene expression results. The PPC consists of a predispensed synthetic DNA template and a primer set that can amplify it. The sequence of the DNA template is not present in the human genome, and thus tests the efficiency of the polymerase chain reaction itself.
- The No Template Control (NTC) is strongly recommended, and can be used to monitor the DNA contamination introduced during the workflow such as reagents, tips, and the lab bench.

Kit Components

Component	Cat #	Quantity	Storage
GeneQuery™ array plate with lyophilized primers	GK045	1	4°C or -20°C
Optical PCR plate seal	N/A	1	RT
Nuclease-free H ₂ O	GQ100-1	2	4°C

Additional Materials Required (Materials Not Included in Kit)

Component	Recommended
Reverse transcriptase	First-Strand cDNA Synthesis Master Mix, 4x (ScienCell, Cat #MB6008)
cDNA template	Customers' samples
qPCR master mix	GoldNStart TaqGreen qPCR Master Mix (ScienCell, Cat #MB6018)

Quality Control

All the primer sets are validated by qPCR with melt curve analysis. The PCR products are analyzed by gel electrophoresis. Single band amplification is confirmed for each set of primers.

Product Use

GQH-HST is for research use only. It is not approved for human or animal use, or for application in clinical or *in vitro* diagnostic procedures.

Shipping and Storage

The product is shipped at ambient temperature. Upon receipt, the plate should be stored at 4°C and is good for up to 12 months. For long-term storage (>1 year), store the plate at -20°C in a manual defrost freezer.

Procedures

Note: The primers in each well are lyophilized.

1. Prior to use, allow plates to warm to room temperature.
2. Briefly centrifuge at 1,500x g for 1 minute before slowly peeling off the seal.
3. Prepare 20 μ l PCR reactions for one well as shown in Table 1.

Table 1

cDNA template	0.2 – 250 ng
2x qPCR master mix	10 μ l
Nuclease-free H ₂ O	variable
Total volume	20 μl

Important: Only use polymerases with hot-start capability to prevent possible primer-dimer formation. Only use nuclease-free reagents in PCR amplification.

4. Add the mixture of 2x qPCR master mix, cDNA template, and nuclease-free H₂O to each well containing the lyophilized primers. Seal the plate with the provided optical PCR plate seal.

Important: In NTC control well, do NOT add cDNA template. Add 2x qPCR master mix and nuclease-free H₂O only.

5. Briefly centrifuge the plates at 1,500x g for 1 minute at room temperature. For maximum reliability, replicates are strongly recommended (minimum of 3).
6. For PCR program setup, please refer to the instructions of the master mix of the user's choice. We recommend a typical 3-step qPCR protocol for a 200nt amplicon:

Three-step cycling protocol

Step	Temperature	Time	Number of cycles
Initial denaturation	95°C	10 min	1
Denaturation	95°C	20 sec	40
Annealing	65°C	20 sec	
Extension	72°C	20 sec	
Data acquisition	Plate read		
<i>Recommended</i>	<i>Melting curve analysis</i>		1
Hold	4°C	Indefinite	1

7. (Optional) Load the PCR products on 1.5% agarose gel and perform electrophoresis to confirm the single band amplification in each well.

Figure 1. Layout of GeneQuery™ qPCR array kit controls.

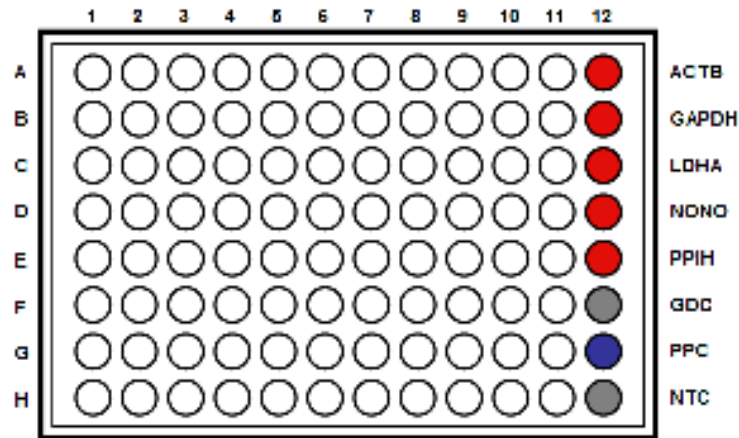


Table 2. Interpretation of control results:

<i>Controls</i>	<i>Results</i>	<i>Interpretation</i>	<i>Suggestions</i>
Housekeeping gene controls	Variability of a housekeeping gene's C _q value	The expression of the housekeeping gene is variable in samples; cycling program is incorrect	Choose a constantly expressed target, or analyze expression levels of multiple housekeeping genes; use correct cycling program and make sure that all cycle parameters have been correctly entered
gDNA Control (GDC)	C _q ≥ 35	No gDNA detected	N/A
	C _q < 35	The sample is contaminated with gDNA	Perform DNase digestion during RNA purification step
Positive PCR Control (PPC)	C _q > 30; or The C _q variations > 2 between qPCR Arrays.	Poor PCR performance; possible PCR inhibitor in reactions; cycling program incorrect	Eliminate inhibitor by purifying samples; use correct cycling program and make sure that all cycle parameters have been correctly entered
No Template Control (NTC)	Positive	DNA contamination in workflow	Eliminate sources of DNA contamination (reagents, plastics, etc.)

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Figure 2. A typical amplification curve showing the amplification of a qPCR product.

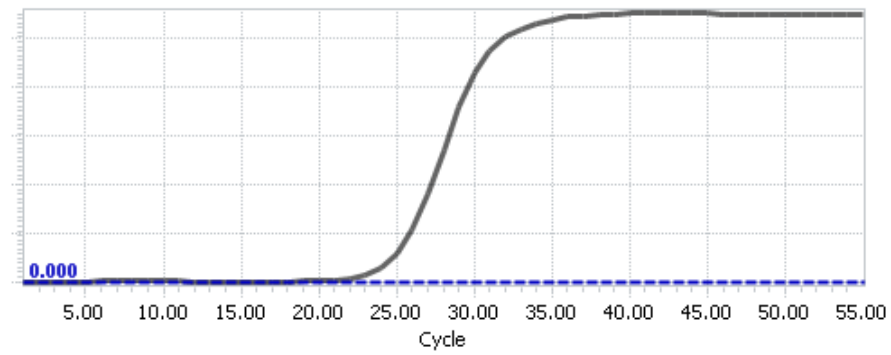
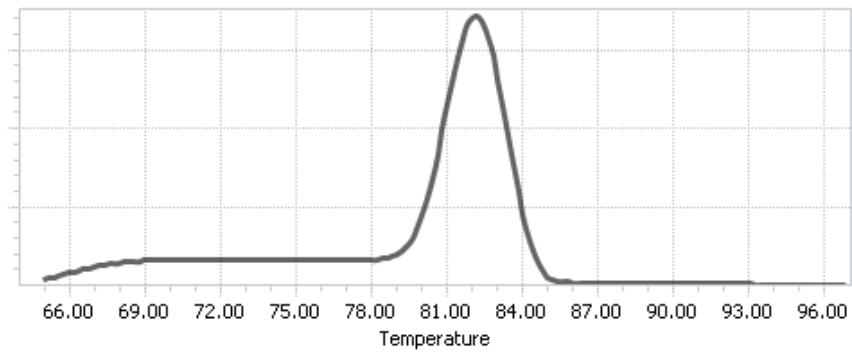


Figure 3. A typical melting peak of a qPCR product.



Quantification Method: Comparative $\Delta\Delta Cq$ (Quantification Cycle Value) Method

1. **Note:** Please refer to your qPCR instrument's data analysis software for data analysis. The method provided here serves as guidance for quick manual calculations.

You can use one or more housekeeping genes as a reference to normalize samples.

Important: We highly recommend using all 5 housekeeping genes included in this kit: ACTB, GAPDH, LDHA, NONO, and PPIH.

2. For a single housekeeping gene, ΔCq (ref) is the quantification cycle number change for that housekeeping gene (HKG) between an experimental sample and control sample.

$$\Delta Cq \text{ (ref)} = Cq \text{ (HKG, experimental sample)} - Cq \text{ (HKG, control sample)}$$

When using multiple housekeeping genes as a reference, we recommend normalizing using the geometric mean [1] of the expression level change, which is the same as normalizing using the arithmetic mean of ΔCq of the selected housekeeping genes.

$\Delta Cq \text{ (ref)} = \text{average} (\Delta Cq \text{ (HKG1)}, \Delta Cq \text{ (HKG2)}, \dots, \Delta Cq \text{ (HKG n)})$ (n is the number of housekeeping genes selected)

If using all 5 housekeeping genes included in this kit (ACTB, GAPDH, LDHA, NONO, and PPIH) use the following formula:

$$\Delta Cq \text{ (ref)} = (\Delta Cq(\text{ACTB}) + \Delta Cq(\text{GAPDH}) + \Delta Cq(\text{LDHA}) + \Delta Cq(\text{NONO}) + \Delta Cq(\text{PPIH})) / 5$$

Note: ΔCq (HKG) = Cq (HKG, experimental sample) - Cq (HKG, control sample), and ΔCq (HKG) value can be positive, 0, or negative.

3. For any of your genes of interest (GOI),

$$\Delta Cq \text{ (GOI)} = Cq \text{ (GOI, experimental sample)} - Cq \text{ (GOI, control sample)}$$

$$\Delta\Delta Cq = \Delta Cq \text{ (GOI)} - \Delta Cq \text{ (ref)}$$

$$\text{Normalized GOI expression level fold change} = 2^{-\Delta\Delta Cq}$$

References

[1] Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paepe A, Speleman F. (2002) "Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes." *Genome Biol.* 3(7): 1-12.

Example: Comparative $\Delta\Delta Cq$ (Quantification Cycle Value) Method

Table 3. Cq (Quantification Cycle) values of 2 genes-of-interest and 5 housekeeping genes obtained for experimental and control samples.

Samples	Genes of Interest		Housekeeping Genes				
	GOI1	GOI2	<i>ACTB</i>	<i>GAPDH</i>	<i>LDHA</i>	<i>NONO</i>	<i>PPIH</i>
Experimental	21.61	22.19	17.16	17.84	20.12	19.64	26.40
Control	33.13	26.47	18.20	18.48	20.57	19.50	26.55

$$\begin{aligned}\Delta Cq(\text{ref}) &= (\Delta Cq(\text{ACTB}) + \Delta Cq(\text{GAPDH}) + \Delta Cq(\text{LDHA}) + \Delta Cq(\text{NONO}) + \Delta Cq(\text{PPIH})) / 5 \\ &= ((17.16 - 18.20) + (17.84 - 18.48) + (20.12 - 20.57) + (19.64 - 19.50) + (26.40 - 26.55)) / 5 \\ &= -0.43\end{aligned}$$

$$\begin{aligned}\Delta Cq(\text{GOI1}) &= 21.61 - 33.13 \\ &= -11.52\end{aligned}$$

$$\begin{aligned}\Delta Cq(\text{GOI2}) &= 22.19 - 26.47 \\ &= -4.28\end{aligned}$$

$$\begin{aligned}\Delta\Delta Cq(\text{GOI1}) &= \Delta Cq(\text{GOI1}) - \Delta Cq(\text{ref}) \\ &= -11.52 - (-0.43) \\ &= -11.09\end{aligned}$$

$$\begin{aligned}\Delta\Delta Cq(\text{GOI2}) &= \Delta Cq(\text{GOI2}) - \Delta Cq(\text{ref}) \\ &= -4.28 - (-0.43) \\ &= -3.85\end{aligned}$$

$$\begin{aligned}\text{Normalized GOI1 expression level fold change} &= 2^{-\Delta\Delta Cq(\text{GOI1})} \\ &= 2^{11.09} \\ &= 2180\end{aligned}$$

$$\begin{aligned}\text{Normalized GOI2 expression level fold change} &= 2^{-\Delta\Delta Cq(\text{GOI2})} \\ &= 2^{3.85} \\ &= 14.4\end{aligned}$$

Conclusion: Upon treatment, expression level of GOI1 increased 2,180 fold, and expression level of GOI2 increased 14.4 fold.

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GeneQuery™ Human Hepatic Steatosis qPCR Array Plate Layout*
(***8 controls*** in Bold and Italic)

	1	2	3	4	5	6	7	8	9	10	11	12
A	ABCA1	ADIPOR2	CES1	FASN	GSK3B	INSR	LPL	OAT	PNPLA3	RBP4	SLC2A4	<i>ACTB</i>
B	ABCB11	AKT1	CNBP	FOXA2	HADHA	IRS1	MAPK1	OTC	PPARA	RXRA	SOCS3	<i>GAPDH</i>
C	ACADL	APOA1	CPT1A	FOXO1	HMGCL	IRS4	MAPK8	PCK2	PPARG	SCD	SQSTM1	<i>LDHA</i>
D	ACADM	APOB	CPT1B	G6PC	HMGCR	KRT18	MLXIPL	PDE3B	PPARGC1A	SERPINE1	SREBF1	<i>NONO</i>
E	ACLY	APOC3	CPT2	G6PD	HSD3B7	LDLR	MTOR	PDK4	PRKAA1	SLC17A5	SREBF2	<i>PPIH</i>
F	ACOX1	APOE	CYP2E1	GFPT1	IGF1	LEP	NFKB1	PIK3CA	PRKCSH	SLC22A5	STAT3	<i>GDC</i>
G	ADIPOQ	ASS1	CYP7A1	GPD1	IGFBP1	LEPR	NR1H3	PIK3R1	PTPN1	SLC2A1	TNF	<i>PPC</i>
H	ADIPOR1	CD36	FABP1	GPT	INS	LMNA	NR1H4	PKLR	PTRF	SLC2A2	XBP1	<i>NTC</i>

* Gene selection may be updated based on new research and development

Appendix. Plate type choice chart.

Plate type A

Brand	Model	kit catalog #
ABI / Life Tech	ABI 5700	GK045-A
	ABI 7000	GK045-A
	ABI 7300	GK045-A
	ABI 7500	GK045-A
	ABI 7700	GK045-A
	ABI 7900 HT	GK045-A
	QuantStudio	GK045-A
	ViiA 7	GK045-A
Bio-Rad	Chromo4	GK045-A
	iCycler	GK045-A
	iQ5	GK045-A
	MyiQ	GK045-A
	MyiQ2	GK045-A
Eppendorf / Life Tech	Matercyler ep realplex 2	GK045-A
	Matercyler ep realplex 4	GK045-A
Stratagene	MX3000P	GK045-A
	MX3005P	GK045-A

Plate type B

Brand	Model	kit catalog #
ABI / Life Tech	ABI 7500 Fast	GK045-B
	ABI 7900 HT Fast	GK045-B
	QuantStudio Fast	GK045-B
	StepOnePlus	GK045-B
	ViiA 7 Fast	GK045-B
Bio-Rad	CFX Connect	GK045-B
	CFX96	GK045-B
	DNA Engine Opticon 2	GK045-B
Stratagene	MX4000	GK045-B

Plate type C

Brand	Model	kit catalog #
Roche	Lightcycler 96	GK045-C
	Lightcycler 480 (96-well)	GK045-C